# Assessing impact of fishing on growth of corkwing wrasse (Symphodus melops) and goldsinny wrasse (Ctenolabrus rupestris) 

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#### Abstract

Corkwing wrasse (Symphodus melops) and goldsinny wrasse (Ctenolabrus rupestris) are two temperate wrasse (Labridae) species who are targeted by commercial fishery due to their ability to reduce sea-lice infections. The scientific community have raised concerns regarding the longterm sustainability of the wrasse fisheries, as these two species are extensively harvested along the Norwegian coast. Studies on a wrasse fishery impact in wild populations of corkwing and goldsinny wrasse are therefore needed. Addressing whether harvesting impacts on the growth rate in these two species can provide a better understanding on how wild populations of wrasse are affected. Giving a better basis for future regulations and directions to maintain a sustainable wrasse fishery.


In this study I analyzed five years of catch-mark-recapture (CMR) data on individual growth collected in a marine protected area (MPA) on the west coast of Norway. A before-after controlimpact (BACI) approach on two adjacent islands (fished and control (MPA)) allowed me to assess the possible effect of a replicated wrasse fishery on growth rates in wild populations of corkwing and goldsinny wrasse. A total of 8855 of corkwing and 4993 of goldsinny were tagged over thirteen sampling periods between 2017 and 2021. In total 1890 corkwing and 733 goldsinny were recaptured. My results showed a significant difference in growth rate between fished area and control area for male corkwing wrasse. Males in the fished area had a faster growth after fishing started compared to before. This could indicate that fishing had a clear effect on male corkwing growth. In the long run, this may be a positive sign for the fished population since males growing larger quicker could lead to earlier maturation, better male parental care, and better population productivity. Female corkwing wrasse did not show significant differences in growth rate between the areas, neither before nor after fishing started. Earlier maturation could be a possible explanation, with more investment towards gonad growth instead of growth. For goldsinny there were little difference in growth between fished and control area, neither before nor after fishing started. A possible explanation could be that both sexes of goldsinny grow slower than corkwing, meaning that it could take longer time to detect any differences in growth. Based on my results, there is reasons to believe that a wrasse fishery will have a greater impact on corkwing compared to goldsinny. As corkwing showed more difference between the two areas. Goldsinny did not have the same growth variation between the areas. This could indicate that fishing can have less negative effect than anticipated, especially if faster growing males provide better parental care.

## Sammendrag

Grønngylt (Symphodus melops) og bergnebb (Ctenolabrus rupestris) er to av tre tempererte leppefisk (Labridae) arter som blir målrettet fisket for å bruke i oppdrettsanlegg, på grunn av deres evne til å begrense lakseluspåslag. Forskningsmiljøet har uttrykt bekymring angående den langsiktige bærekraften til leppefiskeriene, ettersom fisking på disse to artene er omfattende langs norskekysten. Flere studier på hvordan leppefiske påvirket ville populasjoner av grønngylt og bergnebb er derfor nødvendig. Ved å adressere mulige påvirkninger fiskeri har på vekst rate for disse to artene, kan føre til en bedre forståelse for hvordan ville populasjoner av leppefisk blir påvirket. Noe som fører til et bedre fundament for fremtidige reguleringer og direktiver for å opprettholde et bærekraftig leppefiske.

I dette studiet brukte jeg fem år med fangst-merking-gjenfangst data fra et marint verneområde på vestkysten av Norge. På to nærliggende øyer (fisket og kontroll (marint verneområde)) ble det brukt en før-etter tilnærming for å se på effektene av et etterlignet leppefiske på individuell vekstrate i populasjoner av grønngylt og bergnebb. Totalt 8855 grønngylt og 4993 bergnebb ble merket over tretten perioder mellom 2017 og 2021. Hvorav totalt 1890 grønngylt og 733 bergnebb ble fanget om igjen. Mine resultater indikerte at det var en signifikant forskjell i vekst rate mellom fisket og kontroll område for grønngylt hanner. Hanner i fisket område hadde raskere vekst etter at fisking startet sammenlignet med før fisking. Dette kan indikere at fisking hadde en klar effekt på vekst for grønngylt hanner. Lenger fremme i tid kan dette være et positivt tegn for den fiskede populasjonen ettersom raskere vekst for hanner kan føre til tidligere modning, bedre foreldreomsorg, og bedre produktivitet i populasjonen. Grønngylt hunner hadde ikke en signifikant forskjell i vekst rate mellom områdene, og heller ikke før eller etter fisking startet. Tidligere modning kan være en mulig forklaring, med mer investering i gonadevekst istedenfor vekst. For bergnebb var lite forskjell mellom vekst i fisket område og kontroll område og heller ikke før og etter fisking startet. En mulig forklaring kan være at begge kjønnene vokser saktere enn grønngylt, som betyr at det kan ta lenger tid å se forskjeller i vekst. Basert på mine resultater, så er det grunner til å tro at leppefiske vil ha en større effekt på grønngylt enn bergnebb. Ettersom grønngylt viste mer variasjon i vekst mellom de to områdene, med klare forskjeller mellom kjønnene. Bergnebb hadde ikke den samme variasjonen i vekst mellom områdene. Forvaltning av disse artene kan by på utfordringer ettersom de blir fisket opp i store mengder for å bli brukt som rensefisk i lakseoppdrett.

## Preface

First, I rd like to express my deepest thanks to my supervisors Tonje Knutsen Sørdalen and Kim Halvorsen for their incredible support, guidance, and feedback throughout the project. I would not have made it without you and could have not asked for better supervisors! Your knowledge and positivity truly made this project inspiring and fun to work with. I would also like to thank Kim Halvorsen and Torkel Larsen for sharing their experience and helping me in field work with identifications of wrasse, throwing and hauling fyke nets. In addition, I would also thank Reidun Bjelland and Anne Berit Skiftesvik on the Institute of Marine Research on Austevoll for their contribution on data sampling in the field work. I'd also like to thank previous contributors from earlier sampling years for their collection of valuable data.

Thank you to my wonderful girlfriend who also wrote her master thesis at the same time. I am truly grateful for your support and that we could share our ups and downs together. Lastly, I'd like to thank my family for their consistent support and kind words.
<Kristiansand 20.05.2022>
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## 1. Introduction

### 1.1 Marine fisheries and its impacts

Commercially and recreationally fisheries are important to local and national economies, and contributes to substantial economic benefits (Arlinghaus et al., 2002; Hilborn et al., 2003). However, the sustainability of fisheries has been questioned, and remains as a major problem worldwide (Sparholt et al., 2021). Overfishing of the spawning biomass of a population, recruitment overfishing, can reduce its ability to replenish and can consequently lead to population collapse (Pauly, 1994; Ben-Hasan et al., 2021) which the collapse of north western Atlantic Cod (Gadus morhua) is a good example of (Walters and Maguire, 1996).

Although fish stocks may adapt to fishing through evolution in the long run, fishing operations are still deliberately selective_(Pauli et al., 2015) not only as a result of regulations enacted to protect smaller individuals, but also by fishermen targeting commercially profitable and available species (Salas et al., 2004; Andersen et al., 2012). When it comes to selectivity, population productivity may negatively be affected by the size selectivity (i.e., removal of large individuals) and mortality that is imposed by many commercial and recreational fisheries (UusiHeikkilä et al., 2015b). Earlier studies have suggested that smaller individuals (subjected to low fishing mortality) will typically have low fecundity and reproductive success (Shelton, 2006; Uusi-Heikkilä et al., 2015a).Large males that are more dominant in male-male competition and have a high resource-holding potential are often preferred by the large females (van den Berghe and Gross, 1989; Sørdalen et al., 2018). In systems where body size is a sexually selected trait, a reduction in size variability can be expected to disrupt the choice of mate and competition within the species. Given that a size variability is present in a exploited population as a result of size-selective harvesting (Hutchings and Baum, 2005; Nusslé et al., 2017).

Changes in abundance and size structure because of fisheries can have indirect (and often negative) effects on other ecosystem components. Removal of predators high in the food web can completely restructuring the food web in ecosystems, and possibly resulting in cascading effects through the trophic levels below (Frank et al., 2005; Norderhaug and Moland, 2021), ultimately affecting nontargeted species (Wood et al., 2018; Perälä and Kuparinen, 2020). In a longer timeframe, fish stocks may adapt to fishing through evolution (Roos et al., 2006). Most typically, overexploiting harvesting practises tend to select for earlier maturation at smaller size causing higher reproductive investment for the individuals in the exploited stock (Reznick et
al., 1990; Conover and Munch, 2002; Hutchings and Baum, 2005; De Roos et al., 2006; Jørgensen et al., 2007; Hočevar and Kuparinen, 2021). Traits connected to maturation timing can change faster than other life-history traits, and therefore may be more responsive to fisheries induced evolution (FIE) (Audzijonyte et al., 2013).

Precautionary approaches to promote rebuilding and limit the risk of fish collapse under sustained fishing pressure have been supplemented with a reference point for management called maximum sustainable yield (MSY) (UNCLOS, 1982). Long-term predictions of yield and sustainability generally assume density dependence in the abundance of incoming recruits by incorporating a negative relationship between recruitment and spawning stock biomass (Cadigan 2013).

### 1.2 Density dependent growth

The removal of fish from a population will potentially ease the competition of space and food, leaving more resources for the remaining individuals in the populations to increase their growth rate. This concept in population ecology is called density dependent ,and occurs when population growth rates or survival are regulated by the density of a population (Ricker, 1954; Beverton and Holt, 2012). When the number of individuals in a population becomes high, the mortality could increase with higher intra-specific competition for limited resources such as food and habitat. In addition, diseases, parasites and predators are other mechanisms for density dependent mortality (Hixon and Webster, 2002; Stige et al., 2019). High density would also increase competition for spawning territories and mates, which could reduce fertility (Anderson and May, 1978; Pulliam, 1988; Sinclair et al., 2003). The opposite would be the case when population size is low. Mortality rates during the juvenile (pre-recruit) phase is typically very high and predation is thought to be the main cause (Cushing, 1974; Sogard, 1992). Therefore, it is believed that fish populations are mainly regulated by density-dependent growth in the juvenile phase (Lorenzen and Enberg, 2002). Increase of food availability due to low competition could therefore be particularly advantageous for juvenile individuals that are dependent on rapid growth in an early, vulnerable life stage (Andersen et al., 2017). Rapid growth makes it possible for a newly settled juvenile to leave the most vulnerable size classes quicker, resulting in an advantage over slower growing individuals (Sogard, 1992). Density regulations can be challenging to detect since populations can fluctuate widely, without showing signs of density regulations, this has caused debate regarding how regulations can be
detected and whether it is important at all (Turchin, 1995; Hixon and Webster, 2002; Brook and Bradshaw, 2006). Fish growth is believed to be an important key in regulating fish populations (Lorenzen and Enberg, 2002). Fish species tend to grow indeterminately (somatic growth), with very few exceptions, which means fish grow their whole life. This somatic growth can have an impact on survival, sexual maturity and productiveness (Rose et al., 2001; Vincenzi et al., 2014; Korman et al., 2021). Some strategies to avoid juvenile mortality are rapid growth and early sexual maturity, which is expected to give better fitness and incur predation risks (Bacon et al., 2005).

### 1.3 Marine protected areas

The density of populations can be increased by conservation efforts, such as establishing notake areas or marine protected areas (MPAs), areas partially or completely closed for fishing. It is documented that MPAs create areas with higher fish biomass and size distributions compared to unprotected fished areas, if designed correctly (Halpern et al., 2010; Edgar et al., 2014; Baskett and Barnett, 2015). Support from fishers is still limited, even though MPA advocates suggests that these protected areas not only protect the exploited species, but also provides "spillover" effects where fish or larvae leave the MPA and are eventually caught (McClanahan and Mangi, 2000; Hilborn, 2018). Hoping that spillover from a high biomass area within a MPA can offset the assumed loss of catch associated with their establishment (Gell and Roberts, 2003; Grorud-Colvert et al., 2014; Marshall et al., 2019).

For the fished areas, there is reasonable to believe that removal of individuals will have a positive effect on population growth, as more food resources are distributed on fewer individuals in the population. However, the opposite may be the case inside the MPA as fish abundance can affect population growth in a negative way, having more competition on limited food resources, leading to overall smaller individuals with slower growth rates over time (Gårdmark et al., 1999; Post et al., 1999; Lorenzen and Enberg, 2002; Beverton and Holt, 2012). It therefore has been hypothesized that MPAs can lead to slowed growth of exploited species, reducing its usefulness as a fisheries management tool (Claudet et al., 2006; Gårdmark et al., 2006; Marshall et al., 2019).

### 1.4 Wrasse fishery

MPAs are well suited to protect and manage sedentary fish and crustacean species in shallow waters (Knutsen et al., 2022). In Norway, small MPAs have been proven to increase size and abundance of two species of wrasse, the corkwing wrasse (Symphodus melops) and goldsinny wrasse (Ctenolabrus rupestris) (Halvorsen et al., 2017a). These two species have become commercially important in Scandinavia and on the British Isles, where they are used as cleaner fish in Salmon Aquaculture (Halvorsen et al., 2021a). Since starting farming on Atlantic salmon (Salmo salar Linnaeus, 1758) in Norway in the 1970`s, removal of ectoparasitic salmon lice (Lepeophteirus salmonis Krøyer, 1837 and Caligus elongatus Nordmann, 1832) have been challenging (Heuch et al., 2005). In addition to economic losses, the salmon lice also cause damage and stress when attached to the skin of the fish and could potentially cause a threat to nearby ecosystems (Costello, 2006; Krkošek et al., 2006). After Bjordal (1988) described the symbiosis between Atlantic salmon and wrasse (Labridae Cuvier, 1816), harvesting of wrasse as a biological delousing tool in the aquaculture industry started (Espeland et al., 2010). The demand for wild caught wrasse increased rapidly in the late 2000`s as salmon lice developed resilience towards commonly used chemical pesticides (Besnier et al., 2014; Skiftesvik et al., 2014). Mechanical and thermal delousing methods are now largely replacing chemical pesticides in Norway. However, both farmed and wild caught wrasse are still extensively used, with corkwing and goldsinny as the most commonly used wild cleaner fish (Faust et al., 2018; Overton et al., 2019). Several limitations have been imposed in order to avoid overfishing, currently the fishery is managed by seasonal closure from mid-July to mid-October, minimum species-specific size limits; 11 cm for goldsinny, rock cook (Centrolabrus exoletus) and cuckoo (Labrus mixtus), 12 cm for corkwing and between $22-28 \mathrm{~cm}$ for ballan wrasse (Labrus bergylta)(Forskrift om regulering av fisket etter leppefisk i 2022, 2021.§8). To avoid overfishing it is important to monitor fisheries and investigate how local populations may be affected by fishing pressure, since depletion of local populations can occur (Halvorsen et al., 2017b).

In this study, I explored the impact of fishing on adult growth in two wrasse species. Of these, sexual size dimorphism is strongly male-biased in the corkwing wrasse, while male and females goldsinny wrasse are more similar. In many species the form and strength of selection on body size or correlated traits is rarely identical between the sexes, resulting in sexual size dimorphism (SSD) (Parker, 1992; Fairbairn et al., 2007; Halvorsen et al., 2016). Nesting males of corkwing
wrasse is a good example, as they are fiercely territorial during nesting season, leading to aggressive confrontations, often won by large individuals (Potts, 1974a, 1985). For different reasons, sexual size dimorphism is often overlooked when assessing the management of commercial fisheries. Size selective harvesting on sexually dimorphic populations would likely be sex-selective, which makes the neglection in the management assessment unfortunate(Rowe and Hutchings, 2003; Fenberg and Roy, 2008; Hanson et al., 2008; Zhou et al., 2010; Kendall and Quinn, 2013; Halvorsen et al., 2016). Moreover, the corkwing wrasse grow generally faster and has shorter life span than the goldsinny (Halvorsen et al., 2017a). Thus, the growth response to fishing may differ between and within these two species.

The objective with this study was to compare growth in corkwing and goldsinny wrasse populations between a MPA and an adjacent fished area. Based on five years of capture-markrecapture (CMR) data from 13 sampling periods, I use linear models to investigate if a simulated wrasse fishery has an impact on body growth rates. The models for corkwing are separated between females and males because of the known differences in growth between the sexes. Goldsinny males and females were presented in the same model because both sexes have similar growth rates and fewer CMR data. I hypothesize that corkwing individuals in the fished area will have faster growth rate based on theories suggesting there will be more space and food available for the remaining individuals after larger fish is removed due to fishing. For goldsinny I hypothesize that the results will present less difference in growth rates and fishing will have a smaller impact compared to corkwing. I imagine goldsinny are more effected by other local factors such as density, predation, and habitat.

## 2. Methods

### 2.1. Study area

The study was conducted inside an experimental marine protected area (MPA) nearby the field station of Institute of Marine Research in Austevoll, outside of Bergen in Western Norway. Three islands are located within the MPA (Fig 1) where commercial fishing for wrasse has been prohibited since 2017. The surrounding waters of two of the islands, Bleikjo and Saltskjærholmane, was used as the study area. With the southeastern side of the islands facing Huftarøy being more sheltered to Bjørnafjorden compared to the east side. The two islands were divided into multiple zones. Bleikjo into 4 zones (1-4) and Saltskjærholmane into 12 zones (1-12) (Aasen, 2019). At low tide the average length of the shoreline at each zone is 79.8 at Bleikjo and 141.3 at Saltskjærholamen. The distance between the islands are 80 meters and with a maximum water depth of about 25 meters. The surrounding habitats mostly consist of hardbottom covered by a variety of kelp, which is a suitable habitat for wrasse fishes (Skiftesvik et al., 2014). In a before-after control-impact (BACI) approach the smallest island of the two, Bleikjo, was chosen as the site for the experimental fishery, called fished area from now on. Saltskjærholmane, the unfished site, will be referred to as control area.


Figure 1: Overview map of the geographic positions of Bleikjo and Saltskjorholmane, at Austevoll in Hordaland. Location in Norway are marked with a red dot in the top right corner. Saltskjarholmane, Bleikjo and Lambøya/Lambøyskjaret are all a part of the MPA. Map are obtained from https://kart.fiskeridir.no/fiskeri and https://www.norgeskart.no

### 2.2. BACI-design

Before-after control-impact (BACI) are suggested to be statistical powerful designs in environmental impacts assessment studies (Smokorowski and Randall, 2017). Even in the early days of marine conservation science discipline, the BACI design was suggested (Jones et al., 1993). An important tool in environmental impact assessment is to detect changes in a site before the impact happened and after, and compare this to a control location, where the impact persists. A BACI approach will give the unequivocal detection of change between these sites, which strengthens this approach (Moland et al., 2021). Wrasse harvesting in an ecosystem is an example of an anthropogenic disturbance, in a marine conservation setting. Removal of one or part of these anthropogenic disturbances in this setting will represented the "impact".

### 2.3. Study species

Corkwing and goldsinny both belong to the family Labridae which includes more than 500 described species worldwide (Parenti and Randall, 2000; Jansson et al., 2020). These are two of six wrasse species inhabiting the Norwegian coast and are the two most abundant cleaner wrasse species used in Atlantic salmon farming industry in Norway (Blanco Gonzalez and de Boer, 2017). Corkwing functions as intermediate predators in the ecosystem and prey on different invertebrates (Helfman et al., 2009; Skiftesvik et al., 2014). In turn, they are preyed upon by other larger fishes and piscivorous birds (Svåsand et al., 2000; Nedreaas et al., 2008; Dehnhard et al., 2021)Corkwing can reach sizes up to 200-300 millimeters and prefer to inhabit shallow coastal areas (<5 meters depth) filled with a rocks and eelgrass, however they can occur in depths of 15-18 meters as well. Algae is important in corkwing habitats, both as a nest building material and habitats for small crustaceans, gastropods, and bivalves which corkwing feed on (Potts, 1974a, 1985; Costello, 1991a; Sayer et al., 1996). Therefore, they are typically found in areas with high algal cover, e.g., kelp forests and eelgrass beds, within the 5 meters depth (Quignard and Pras, 1986; Gibson, 1993; Skiftesvik et al., 2014). The spawning period of both corkwing and goldsinny starts in May and end late July, with June as the most active month (Costello, 1991a; Skiftesvik et al., 2015).


Figure 2: The wrasses of Norway. Upper left: Ballan wrasse. Upper left: Corkwing wrasse, larger male in the back and female in the front. Center: Rock cook wrasse, females are similar, but less blue. Bottom left: Cuckoo wrasse, red female in the front and blue male in the back. Bottom center: the rarer Scale-rayed wrasse. Bottom right: Goldsinny wrasse. Illustration by Stein Mortensen. Modified with English names and gender signs, with permission from Stein Mortensen

During sexual maturation, corkwing undergoes morphological changes with strong coloration in males that lasts through the spawning season. (Potts, 1974a). Reproductive corkwing males are dimorphic, with one male morph being distinctively blue and green colored, and one male morph employing female mimics (sneaker male). Sneaker males have less distinctive colors and possesses a papilla which resembles that of females. This imitation and alternative reproductive tactic (ART) may increase the chance to be tolerated by territorial dominant males of the same species, leading to a higher likelihood for participating in matings initiated by these males(Gross, 1982; van den Berghe and Gross, 1989; Uglem et al., 2000; Uglem and Rosenqvist, 2002). On average, most corkwing matures when reaching 2-3 years, with a mean length of 100 mm . However, size and age at which fish mature can vary between populations (Potts, 1974b; Costello, 1991b; Darwall et al., 1992; Halvorsen et al., 2016). Corkwing can attain a total length of 28 cm and maximum age of nine years (Darwall et al., 1992; Sayer et al., 1996). While goldsinny may reach a body length of 18 cm and 20 years of age (Darwall et al., 1992; Sayer et al., 1995).

Goldsinny inhabits the same inshore, rocky, and algal habitats as corkwing (Hilldén, 1981; Sundt and Jørstad, 1998; Jansson et al., 2020). However, it is assumed that corkwing in general prefers deeper water and are the least abundant of the two (Halvorsen et al., 2020). Females and males can often be visually distinguishable, as females have rib shaped patterns and males may have orange horizontal stripes on the lower part of their abdomen (Hilldén, 1981). While the nesting corkwing males are brightly colored in green, blue, and orange, the females are plainer with mostly a yellowish or brown color, and also a dark urogenital papilla (Potts, 1974a). Goldsinny males have a red coloration on the abdomen, which separates them from the females (Hilldén, 1981).

### 2.4 Data collection

Data from thirteen sampling periods spread over 5 years of sampling was used to calculate growth (Table 1). For sampling of fish, we used fyke nets with 7.8 m single leader, 70 cm entrance ring and leader mesh size of 11 mm . During a period of six days, eight fyke nets were placed out per day. The fyke nets were placed out in the afternoon and left overnight, which resulted in a soak time between 15-20 hours approximately. Placement of the fyke nets alternated between the zones ( $n=16$ ), each zone was sampled for a total of three times every other day per sampling period. Fyke nets were placed near the shoreline in a perpendicular position, attempting not to place it too deep. The leader net extended towards the surface which would lead most of the passing individuals that move along the bottom into the chambers of the fyke net.

Table 1: Overview of the sampling periods for the standardized fyke-net survey. 2017 had three smaller sampling periods.

| Year |  | Period |
| :---: | :---: | :---: |
| 2018 | 1 | Date (dd.mm/yy) |
|  | 2 | $02.08-08.09 / 2017$ |
| 2019 | 3 | $11.05-18.05 / 2018$ |
|  | 4 | $02.07-09.07 / 2018$ |
|  | 5 | $04.09-11.09 / 2018$ |
| 2020 | 6 | $09.05-23.05 / 2019$ |
|  | 7 | $03.09-08.09 / 2019$ |
|  | 8 | $04.05-09.05 / 2020$ |
| 2021 | 9 | $27.06-02.07 / 2020$ |
|  | 10 | $26.08-31.08 / 2020$ |
|  | 11 | $06.05-11.05 / 2021$ |
|  | 12 | $28.06-03.07 / 2021$ |
|  | 13 | $23.08-28.08 / 2021$ |
|  |  |  |

Individuals caught was determined to species level and measured for total length to the nearest millimeter. Further, light pressure was applied on the abdomen to gather information about the sex and spawning state (female, male or sneaker), based on sexual products extruded. When there were no sexual products present, sex was determined by looking at phenotype for goldsinny and corkwing wrasse. Sneaker males were only distinguishable in the mating season when sexual products where extruded, because of their female phenotype. After each fyke, we gently released all catch at the same location as capture.

### 2.5 PIT-tagging

Passive Integrated Transponder (PIT) tags were used to monitor the fate of the wrasse individuals. These are glass-encapsuled passive transponders that are sealed in biocompatible glass to protect the electronics and prevent tissue irritation (Gibbons and Andrews, 2004). For corkwing, PIT-tags have been used previously to estimate fishing mortality (Halvorsen et al., 2017c), and the use of PIT-tags are well-documented for wrasse and as an identification method for studies on fish (PRENTICE and F., 1990; Bolland et al., 2009).
For this study, we used half duplex PIT-tags ( $2,12 \times 12 \mathrm{~mm}$; RFID Solutions Stavanger, Norway). Every wrasse species in the catch were scanned (HPR lite-from Biomark Inc. USA) to check for presence of a PIT-tag from earlier tagging periods. For previously tagged individuals, PIT-tag ID numbers were also noted in addition to the method described above. Individuals of ballan, goldsinny and corkwing wrasse $>100 \mathrm{~mm}$ that were not previously tagged were anesthetized prior to tagging in $50-100 \mathrm{mg} 1-1$ tricaine methanesulfonate (MS-222) in $8-101$ of seawater until loss of equilibrium. The loss of equilibrium occurred differently depending on size and species, most of the individuals were ready for tagging within 1-3 minutes. A tag injector with a needle fitted were used to inject a PIT-tag into the body cavity. Between each tag injection, the tag injector was cleaned in $96 \%$ ethanol and the needle was replaced after tagging approximately 20-40 individuals. After tagging, scales were collected from corkwing with a tweezer and put into an empty Eppendorf tube for further processing. Fin clippings on the caudal fin were_collected by using a small scissor for future DNA analysis.

### 2.6 Data analysis

Statistical analyses were carried out using the R software, version 4.0.2 (R Core Team, 2018). The ggplot2 package was used to create all graphics (Wickham, 2016). Linear models were used to test for fishing impact on individual growth. All data was modelled with a gaussian error distribution using the function $\operatorname{lm}()$. A series of assumptions such as independence, normality, homogeneity, fixed X, and correct model specifications ((Zuur et al., 2009). A model validation process is necessary when applying a linear model to your data, to verify these assumptions. By following instructions I: 1) plotted the residuals vs fitted values to check for homogeneity of variance, 2) plotted a QQ plot to check for normality, and 3) plotted the residuals against each explanatory variable to check for independence (Zuur et al., 2009). The growth models were fitted separately for each species and corkwing was also divided between females and males. The response variable in the models included growth (mm per month),
length (mm), and CPUE (number of individuals caught in fyke net). Area (fished and control) and sampling year are the explanatory variables.

To detect if there is an impact of fishing on growth a significant interaction between area and year is needed. The reason for this is that I assume that there is a difference between areas (fished, control). However, to conclude that the difference in growth is directly related to fishing, a significant interaction between area and year is needed. I therefore chose two models including one with an interaction effect and one with only an additive effect: A likelihood ratio test (LRT) was used on each growth model for corkwing female, corkwing male and goldsinny.

$$
\begin{aligned}
& \text { Growth rate }=\text { Previous length }+ \text { Area }+ \text { Year } \\
& \text { Growth rate }=\text { Previous length }+ \text { Area } * \text { Year }
\end{aligned}
$$

## 3. Results

### 3.1. Overview

Overall, 16471 individuals of corkwing ( $\mathrm{n}=10745$ ) and goldsinny $(\mathrm{n}=5726)$ was captured (>100mm). From these, 8855 corkwing and 4993 goldsinny was tagged with a total recapture of 1890 and 733 individuals respectively.

Table 2: Total number of tagged individuals of corkwing and goldsinny between the last tagging period every year (fall) and the following spring + summer following year.

| Sampling intervals | Fall-Spring | Fall-Summer | Total (Fall - Spring + Summer |
| :---: | :---: | :---: | :---: |
| Corkwing |  |  |  |
| $2017-2018$ | 89 | 58 | $\mathbf{1 4 7}$ |
| $2018-2019$ | 64 | 27 | $\mathbf{9 1}$ |
| $2019-2020$ | 20 | 59 | $\mathbf{7 9}$ |
| $2020-2021$ | 38 | 41 | $\mathbf{7 9}$ |
|  |  |  |  |
| Goldsinny |  |  | $\mathbf{3 3}$ |
| $2017-2018$ | 20 | 13 | $\mathbf{3 7}$ |
| $2018-2019$ | 21 | 16 | $\mathbf{2 7}$ |
| $2019-2020$ | 10 | 17 | $\mathbf{3 0}$ |

### 3.2. Mean CPUE and length

Mean CPUE indicates that there were some variations for both goldsinny wrasse (2.4-12.5) and corkwing wrasse (4.9-17.5) between 2018 and 2021 (Fig 3). Goldsinny indicated relatively similar CPUE between the areas, with a high top in September of 2018 and a smaller one in September of 2020. Corkwing had overall higher CPUE compared to goldsinny and indicated that there was more variation in CPUE between the sampling periods, with the highest value from the fished area in May of 2019. There seems to be a trend with higher CPUE values in the fished area compared to control for both species before fishing started. After fishing, the control area seems to have a higher CPUE than fished area for both species. In addition, overall CPUE values for goldsinny seems to follow each other more compared to corkwing.


Figure 3: Mean CPUE for tagged individuals of corkwing and goldsinny in the control and fished area between the first period in 2018 and last period in 2021. Black vertical line indicates when the replicated wrasse fishery on Bleikjo was started, August 2019. Grey vertical lines highlight the continuation of harvesting in the fished area in August every year until 2021.

Findings showed some trends with variations in mean length for corkwing ( $141-147 \mathrm{~mm}$ ) and goldsinny (115.6-119mm) between area and year (Fig 4). Corkwing indicated a relatively stable mean length for fished and control area from 2017 to 2018, with slightly higher mean length in control area. While goldsinny had more variations, with a lower mean length in control overall. The mean length also seemed to increase after fishing was started in the fished area, which could indicate a possible effect of fishing. The opposite was for goldsinny in control area, which resulted in decreasing mean length after fishing started.


Figure 4: Mean length for individuals of corkwing and goldsinny in fished and control area over all sampling years. Year markers are placed in the last sampling period for each year. Black vertical line shows when the replicated wrasse fishery was started on Bleikjo, August 2019.

### 3.3. Growth rate analysis

### 3.3.1. Corkwing

For female corkwing, there were no significant differences in between fished and control area, neither before nor after the fishery ( lm ; area x year effect, likelihood ratio test $\mathrm{L}=-16.165$, d.f. $=7, \mathrm{P}=0.4369$ ) (Fig 5, table 3). Growth rates for female corkwing indicated that the smallest fish had the highest growth rate, before growth declined with increased length. There were some variations in growth rate between the sampling years, with a span around $0-2 \mathrm{~mm}$ monthly overall. Length varied between 100 (minimum size limit for tagging) and 180 mm . The recaptured individuals caught in the tagging periods in 2019 and 2020 had an overall higher estimated growth per month. The lowest growth was observed in 2020-2021, approximately three years after the first experimental fishing was conducted for the first time in the fished area.


Figure 5: Estimated growth per month (mm) of recaptured female corkwing individuals from both areas in the survey. Figure are based on the total number of recaps for every tagging period (May, July, and September) between 2018 and 2021, where every point represents a recaptured individual. Shaded areas present the upper and lower confidence interval for the estimated growth. Before refers to the two first sampling periods before the simulated fishery was started. After refers to the two periods after.

Table 3: Summary of linear model on female corkwing growth rates between year and area. The table shows response variable, coefficients, estimate, standard error (SE), T value an P value. Significant terms are illustrated with a p-value in bold.

| Response | Coefficients | Estimate | SE | T value | P value |
| :---: | :---: | :---: | :---: | :---: | :---: |
| Growth | (Intercept) | 1.753 | 0.279 | 6.290 | $<\mathbf{0 . 0 0 0 1}$ |
|  | length at capture | -0.006 | 0.001 | -4.533 | $<\mathbf{0 . 0 0 0 1}$ |
|  | year 2019 | 0.139 | 0.211 | 0.657 | $\mathbf{0 . 5 1 2}$ |
|  | year 2020 | -0.069 | 0.243 | -0.283 | $\mathbf{0 . 7 7 8}$ |
|  | year 2021 | -0.393 | 0.222 | -1.772 | $\mathbf{0 . 0 7 9}$ |
|  | area (control) | -0.180 | 0.203 | -0.887 | $\mathbf{0 . 3 7 8}$ |
|  | year 2019: area (control) | 0.024 | 0.222 | 0.108 | $\mathbf{0 . 9 1 4}$ |
|  | year 2020: area (control) | 0.310 | 0.262 | 1.186 | $\mathbf{0 . 2 3 8}$ |
|  | year 2021: area (control) | 0.076 | 0.233 | 0.327 | $\mathbf{0 . 7 4 4}$ |

For male corkwing there were a significant difference between the fished and control area, from before and after the fishery (lm; area $x$ year effect, likelihood ratio test $\mathrm{L}=-130.83$, d.f. $=7$, $\mathrm{P}<0.001$ ) (Fig 6, table 4). In the fished area, male corkwing grew slower before fishing and faster after fishing (started August 2019). These interesting results could indicate that fishing had a positive effect on male corkwing growth rate in the fished area. In addition, male corkwing showed similar trends as female corkwing with highest growth for smaller fish, declining with increased length. The growth per month span from around 0 mm to just below 4 mm monthly overall. Length varied between $100-190 \mathrm{~mm}$.


Figure 6: Estimated growth per month (mm) of recaptured male corkwing individuals from both fished and control area. Figure are based on the total number of recaps for every tagging period (May, July, and September) between 2018 and 2021, where every point represents a recaptured individual. Shaded areas present the upper and lower confidence interval for the estimated growth. Before refers to the two first sampling periods before the simulated fishery was started. After refers to the two periods after

Table 4: Summary of linear model on male corkwing growth rates between year and area. The table shows response variable, coefficients, estimate, standard error (SE), T value an P value. Significant terms are illustrated with a p-value in bold.

| Response | Coefficients | Estimate | SE | T value | P value |
| :--- | :---: | :---: | :---: | :---: | :---: |
| Growth | (Intercept) | 2.920 | 0.262 | 11.163 | $<\mathbf{0 . 0 0 0 1}$ |
|  | length at capture | -0.014 | 0.002 | -7.620 | $<\mathbf{0 . 0 0 0 1}$ |
|  | year 2019 | 0.966 | 0.226 | 4.281 | $<\mathbf{0 . 0 0 0 1}$ |
|  | year 2020 | 0.828 | 0.189 | 4.400 | $<\mathbf{0 . 0 0 0 1}$ |
|  | year 2021 | 0.551 | 0.190 | 2.896 | $\mathbf{0 . 0 0 4}$ |
|  | area (control) | 0.441 | 0.160 | 2.755 | $\mathbf{0 . 0 0 7}$ |
|  | year 2019: area (control) | -0.530 | 0.272 | -1.952 | $\mathbf{0 . 0 5 3}$ |
|  | year 2020: area (control) | -0.271 | 0.219 | -1.236 | $\mathbf{0 . 2 1 8}$ |
|  | year 2021: area (control) | -0.978 | 0.229 | -4.284 | $<\mathbf{0 . 0 0 0 1}$ |

### 3.3.2. Goldsinny

As for corkwing, goldsinny did not show any significant difference in growth before or after fishery in the fished and control area ( lm ; area $x$ year effect, likelihood ratio test $\mathrm{L}=31,70$, d.f. $=7, \mathrm{P}=0.4774$ ) (Fig 7, table 5). Individuals had also faster growth for smaller fish, which declined at increased length. Length varied between $100-130 \mathrm{~mm}$. Overall, the growth per month was lower for goldsinny compared to corkwing, with approximately $0-0.6 \mathrm{~mm}$ per month, independent of year, areas, and length. Results indicated relatively similar growth rates between sampling periods from 2017 to 2020 in both areas, where 2017-2018 and 2018-2019 was sampling periods before simulated wrasse fishery started. 2020-2021 had the slowest growth rates, but also resulted in many zero values in growth, that may explain the reason for this.

Goldsinny


Figure 7: Estimated growth per month (mm) of recaptured goldsinny individuals (males and females) from both areas in the survey. Figure are based on the total number of recaps for every tagging period (May, July and September) between 2018 and 2021, where every point represents a recaptured individual. Shaded areas present the upper and lower confidence interval for the estimated growth. Before refers to the two first sampling periods before the simulated fishery was started. After refers to the two periods after

Table 5: Summary of linear model on goldsinny wrasse growth rates between year and area. The table shows response variable, coefficients, estimate, standard error (SE), T value an P value. Significant terms are illustrated with a p-value in bold.

| Response | Coefficients | Estimate | SE | T value | P value |
| :--- | :---: | :---: | :---: | :---: | :---: |
| Growth | (Intercept) | 0.767 | 0.320 | 2.394 | $\mathbf{0 . 0 2 1}$ |
|  | length at capture | -0.005 | 0.003 | -1.924 | $\mathbf{0 . 0 6}$ |
|  | year 2019 | -0.032 | 0.086 | -0.369 | $\mathbf{0 . 7 1 4}$ |
|  | year 2020 | 0.150 | 0.121 | 1.237 | $\mathbf{0 . 2 2 3}$ |
|  | year 2021 | -0.122 | 0.106 | -1.155 | $\mathbf{0 . 2 5 4}$ |
|  | area (control) | -0.053 | 0.084 | -0.626 | $\mathbf{0 . 5 3 4}$ |
|  | year 2019: area (control) | 0.091 | 0.112 | 0.814 | $\mathbf{0 . 4 2 0}$ |
|  | year 2020: area (control) | -0.097 | 0.142 | -0.684 | $\mathbf{0 . 4 9 8}$ |
| year 2021: area (control) | 0.058 | 0.124 | 0.470 | $\mathbf{0 . 6 4 0}$ |  |

## 4. Discussion

The aim of this study was to assess the effects of wrasse fishery on individual growth in wild wrasse populations. By using a before-after control-impact (BACI) approach, and several years of data, I estimated the monthly growth rate for corkwing and goldsinny wrasse from two different sites, a fished area, and inside a MPA established by the IMR (Institute of Marine Research) as control. The main findings were that there was a significant difference in male corkwing growth rate per month between fished and control area. Male corkwing grew faster in the fished area after fishing started, which supports my initial hypothesis where I suggested that corkwing would grow faster in fished area. Therefore, it is reasonable to believe that fishing had a positive effect on this population, with males attaining larger size quicker. The population could benefit from having quick growing males as they provide important parental care for offspring, and removal of the caring sex could have direct consequences for offspring survival (Suski et al., 2003; Sutter et al., 2012). In addition, achieving large sizes quickly can also bring benefits regarding mating opportunities and mate selection by females (Robertson and Hoffman, 1977). Even though my findings suggest that fishing could have a positive effect on male growth rates there are previous studies that suggesting that size selective harvesting of large individuals could reduce body size and age structures within a population (Swain et al., 2007; Fenberg and Roy, 2008). This makes for an interesting contradiction. As fisheries is expected to catch fast growing individuals, but at the same time increase growth for the remaining individuals when density is reduced. Where density dependent seems to affect corkwing males the most. In addition, a previous study in the same MPA did find that the capture probability was negatively correlated with body size (Ruud, 2020). Female corkwing did not show any significant differences in growth rate between the two areas, neither before nor after fishing started. For both areas, estimated growth per month was between 0-2 mm. Females do not provide the same parental care as the males. Investment of more energy towards gonad growth instead of growth and achieving greater reproductive output by earlier maturation could be a possible explanation.

Findings did also indicate clear differences between corkwing sexes, however, it is worth mentioning that this was not tested for in this study. Males grew faster than females, which is also supported by a recent study on factors affecting growth in corkwing in the same area (Vik, 2019). Faster growth for male fish have also been presented in other studies (Treasurer, 1994; Sayer et al., 1995). A possible explanation for slower growth on female corkwing attributed slower female growth to earlier maturation and not the same intrasexual competition as the male-male competition for females will cause (Treasurer, 1994). On the other hand, previous research has suggested that attaining large sizes quickly could be beneficial as large females in general tend to have better fecundity from spawning over longer time periods, higher production of eggs, and create larger sized eggs (Wright and Trippel, 2009; Halvorsen et al., 2016).

For goldsinny sexes were presented in the same model as differences in growth between male and females are negligible. My initial hypothesis was that the smaller-sized goldsinny would have increased growth rate over time, having benefits from being in the fished area where traditional minimum size limit management tool is applied (Halvorsen et al., 2017a). However, my results did not find any significant difference in growth rate between the areas, neither before nor after fishing started. Estimated growth per month for goldsinny was $0-0.6 \mathrm{~mm}$. During modelling, some individuals presented negative growth. Possible explanation for this could be human errors in manually plotting length, and written individual length is shorter compared to previous sampling of the same fish. These had to be corrected to zero growth as the correct length on these individuals are unknown.

In general, smaller fish had the fastest growth per month, and growth gradual declined with length. This result was expected beforehand as high growth rates in smaller individuals can be explained by the vulnerability of being small. Faster growing individuals will be more likely to survive compared to individuals that remain small over a longer time, because slower growth increases the chances for predation(Nilsson and Brönmark, 2000). The reason for that is because most predators are gape-size limited, they are dependent on small enough prey that can fit into their mouths (Post and Parkinson, 2001). Based on adaption strategies among other fish species it is reasonable to believe that corkwing minimize the risk of predation by having high growth rates at small sizes (Tonn et al., 1992; Persson et al., 1996).

The growth predictions for both corkwing wrasse and goldsinny wrasse are based on the hypothesis that growth is density dependent, meaning that growth rates could increase in populations where the largest fish are regularly removed, leaving more food and space available for the remaining fish to grow faster. Density dependent growth have been shown on other fish species where growth opportunities was restricted because of limited food availability and reduced feeding success (Doherty, 1983; Victor, 1986; Jones, 1987; Forrester, 1990; Cowan et al., 2000). Since corkwing and goldsinny have overlapping habitats requirements and depends on the same resources it is reasonable to believe that some sort of resource competition will occur (Costello, 1991a; Sayer et al., 1996; Thangstad, 1999). No fish were removed in the control area, which could lead to a higher density of corkwing and goldsinny competing for the same resources. This might restrict the growth rate over time. Yearly variations in total captures (Appendix A) could be a result of slow growth, inactivity, and low metabolic rates. Studies on ballan wrasse showed that juveniles grow faster at temperatures over $16^{\circ} \mathrm{C}$ (Cavrois-Rogacki et al., 2019). Larger individuals showed low metabolic rates and inactivity at low temperatures $\left(5-10^{\circ} \mathrm{C}\right)$, while physiological performance increased at rising temperatures (Yuen et al., 2019).

Habitat variations will most likely not play an important role in impacting wrasse growth in the two study areas. As a recent study in the same area did not find that variation in habitat influenced corkwing growth. However, the habitats could be too similar to provide differing growth rates (Vik, 2019). On the other hand, previous studies on bluehead wrasse (Thalassa fasciotomy) did find higher growth rates in sheltered inshore populations compared to high exposure populations, claiming that this difference in growth could be a result of different habitat qualities (Warner, 1995). Habitats on the two different islands and zones in this study was generally similar, moderately covered with algae growth with some variation dominating algae type, substrate, and degree of exposure. This has previously been documented as the preferred habitat for wrasse, were corkwing and goldsinny was typically found in sheltered or exposed rocky shores, mudflats and kelp forests, were they feed on slow-moving or sessile prey (Thangstad, 1999; Skiftesvik et al., 2015). The fished area was the smallest and most exposed island of the two and would potentially have different habitats compared to control area, although some zones in the control area was also affected by exposure, mainly by waves. Growth rates should also be expected to be different between the two islands if the nature types have sufficient differences. For instance, places with a higher degree of exposure such as the fished area might provide less vegetation and scree for the fish to use as hiding places in
addition to less available nutrients, compared to the more sheltered control area. However, it might not be as one-sided as this, as corkwing could move between different habitats. Fish movement between the island in this study is limited because of the deeper waters ( $>20 \mathrm{~m}$ ) prevents this (Halvorsen et al., 2021b). A previous study on inshore and offshore populations with many kilometers apart, showed that there was a great variation in exposure between the populations of bluehead wrasse (Schultz and Warner, 1991).

Catch per unit effort (CPUE) indicated little variation between the areas, although there were indications that CPUE had some higher values prior to when the simulated fishery was started in August 2019. Control and fished were the two adjacent islands within the MPA in this study. Estimated catch per unit effort (CPUE) between the two islands showed trends that there are differences between the species, as corkwing have a higher CPUE compared to goldsinny. Variations between sampling periods are also visible, which could be explained by changes in sea temperature in the different sampling periods, affecting fish movement(Deady and Fives, 1995). However, the differences in CPUE between fished area and control area were also relatively similar in 2021, at a point were fish over the minimum size limit ( $>100 \mathrm{~mm}$ ) had been removed annually from the fished area since 2018, when a simulated wrasse fishery started. This result was different compared to a similar study on corkwing and goldsinny (Halvorsen et al., 2017a), which resulted in a higher CPUE for targeted species in MPAs, were corkwing was consistently larger and older inside MPAs. In the same study, goldsinny had less clear growth effects from harvesting. A higher CPUE inside the control area was predicted in advance, as increased density is an anticipated effect of MPAs (McCoy et al., 2010). However, the results showed little differences between the areas. The CPUE estimates was based on tagged fish of corkwing and goldsinny and not total captures, for that reason it is reasonable to believe that limited data samples of these tagged fish could affect the CPUE estimates, even though it may take decades before increased density of a species inside a MPA are detectable (Nickols et al., 2019).

My results indicated that fishing could have an impact on growth in fished populations, and there have been raised concerns on whether and how the Norwegian wrasse fishery impacts the wild populations of wrasse (Espeland et al., 2010; Skiftesvik et al., 2014; Halvorsen et al., 2016). Species-specific size limits, establishments of regional quotas and shorter fishing seasons are some of the recent management regulations (Forskrift om regulering av fisket etter
leppefisk i 2022, 2021. $\S \S 2,4,5,8)$. Still, the Norwegian wrasse fishery is sex- and size selective, with nesting males having higher growth rate, and reaching the minimum size limit before females and sneaker males, which could alter the sex ratios within populations. Previously suggested in corkwing populations on the west coast of Norway, with a strong male-biased dimorphism (Halvorsen et al., 2016, 2017c). Other consequences could be changes in population structure, sex ratios and reproduction (Halvorsen et al., 2016)Reduction and changes in size structure for corkwing and goldsinny populations have previously been connected to wrasse fisheries (Darwall et al., 1992; Sayer et al., 1996). Such depletion of corkwing and goldsinny densities from wrasse fishery may also have a wider consequence on the coastal ecosystems, as removal of wrasse densities can lead to cascade effects through altered predatorprey dynamics (Selden et al., 2017). As a top-down effect mesograzers or as a bottom-up effect as a reduced food source for large piscivores (Kraufvelin et al., 2020; Dehnhard et al., 2021). Wrasse prey on a variety of small grazers in seaweed which could potentially contribute to reduce herbivory/grazing, and hence the community control that small-sized consumer species imposes. In turn, changes in numbers of mesopredatory fish (increase or decrease in abundance) could result in community changes (Norderhaug et al., 2005; Kraufvelin et al., 2020).

Data from this study is based on a catch-mark-recapture (CMR) analysis, which could potentially alter the results if errors occur. The potential for human errors in CMR studies are very much present as catch needs to be handled, tagged, and data are manually recorded in this study. In this study, the negative growth values for goldsinny could be an example of a potential human errors when manually noting the fish length in field. Passive integrated transponders (PIT) are being used to tag individuals, which could potentially lead to tag-loss and/or affect fish survival. Although, usage of PIT is well tested, with minimal chances of tag-loss and little effect on fish`s survival (Peterson et al., 1994; Achord et al., 1996; Gries and Letcher, 2002; Halvorsen et al., 2016). Based on previous testing, I assumed that PIT tags did not affect growth (Kimball and Mace, 2020)

For future research on growth rates for corkwing and goldsinny it would be interesting to continue the CMR-studies in the same area, to see if the fishing impacts on males persist in fished population and maybe affect females and goldsinny as well. Since this study is based on two relatively small islands inside an MPA, it would also be interesting to do more similar studies on other wild wrasse populations in other marine protected areas, to compare with my results. Not only in the western part of Norway, but also in Skagerrak where populations are being harvested and translocated (Skiftesvik et al., 2014; Halvorsen et al., 2017b).

## 5. Concluding remarks

In my thesis, I found a significant difference in growth rate for male corkwing wrasse in the fished area, with males growing faster after fishing was started. This matched with my initial hypothesis, that fish in the fished area would grow faster compared to control area. In addition, this indicated that fishing had an impact on growth rates in the fished population. For female corkwing wrasse there were no significant difference in growth rate between the areas, neither before nor after fishing started. I hypothesized that goldsinny would not have much difference in growth and not be impacted by fishing in the same way as corkwing. This turned out to also be the case, as goldsinny wrasse did not have any significant difference in growth rate between area, neither before nor after fishing. Smaller individuals grew fastest, and corkwing male was found to have the fastest growth per month independent from area. Secondly, the CPUE for corkwing and goldsinny had little variations between the fished are and the control area. Both species showed relatively similar values over several sampling periods.

Lastly, I conclude to say that the impact fishing seemed to have on male corkwing growth in the fished population could have a positive effect on the population. The findings from this study make contributions to future management of wrasse fisheries. As a result of the different impact fishing had on growth rates between males and female corkwing, they should be managed as two different populations in the Western Norway given the difference. Despite that goldsinny did not show any indications of fishing impact on growth rates, it is important with future research on both species to ensure sustainable harvesting.

## References

Aasen, N. L. 2019. The movement of five wrasse species (Labridae) on the Norwegian west coast. http://www.duo.uio.no/.
Achord, S., Matthews, G. M., Johnson, O. W., and Marsh, D. M. 1996. Use of passive integrated transponder (PIT) tags to monitor migration timing of Snake River Chinook salmon smolts. North American Journal of Fisheries Management, 16: 302-313.
Andersen, B. S., Ulrich, C., Eigaard, O. R., and Christensen, A. 2012. me, 69: 131-143.
Andersen, K. H., Jacobsen, N. S., Jansen, T., and Beyer, J. E. 2017. When in life does density dependence occur in fish populations? Fish and Fisheries, 18: 656-667.
Anderson, R. M., and May, R. M. 1978. Regulation and stability of host-parasite population interactions : I . Regulatory processes.: 219-247.
Arlinghaus, R., Mehner, T., and Cowx, I. G. 2002. Reconciling traditional inland fisheries management and sustainability in industrialized countries, with emphasis on Europe. Fish and Fisheries, 3: 261-316.
Audzijonyte, A., Kuparinen, A., and Fulton, E. A. 2013. How fast is fisheries-induced evolution? Quantitative analysis of modelling and empirical studies. Evolutionary Applications, 6: 585-595.
Bacon, P. J., Gurney, W. S. C., Jones, W., Mclaren, I. S., and Youngson, A. F. 2005. Seasonal growth patterns of wild juvenile fish: Partitioning variation among explanatory variables, based on individual growth trajectories of Atlantic salmon (Salmo salar) parr. Journal of Animal Ecology, 74: 1-11.
Baskett, M. L., and Barnett, L. A. 2015. The ecological and evolutionary consequences of marine reserves. Annual Review of Ecology, Evolution, and Systematics, 46: 49-73.
Ben-Hasan, A., Walters, C., Hordyk, A., Christensen, V., and Al-Husaini, M. 2021. Alleviating Growth and Recruitment Overfishing through Simple Management Changes: Insights from an Overexploited Long-Lived Fish. Marine and Coastal Fisheries, 13: 87-98.
Besnier, F., Kent, M., Skern-Mauritzen, R., Lien, S., Malde, K., Edvardsen, R. B., Taylor, S., et al. 2014. Human-induced evolution caught in action: SNP-array reveals rapid amphiatlantic spread of pesticide resistance in the salmon ecotoparasite Lepeophtheirus salmonis. BMC Genomics, 15: 1-18.
Beverton, R. J., and Holt, S. J. 2012. On the dynamics of exploited fish populations. Springer Science \& Business Media, 11.
Bjordal, Å. 1988. Cleaning symbiosis between wrasse (Labridae) and lice infested salmon (Salmo salar) in mariculture. ICES.
Blanco Gonzalez, E., and de Boer, F. 2017. The development of the Norwegian wrasse fishery and the use of wrasses as cleaner fish in the salmon aquaculture industry. Fisheries Science, 83: 661-670. Springer Japan. https://doi.org/10.1007/s12562-017-1110-4.
Bolland, J. D., Cowx, I. G., and Lucas, M. C. 2009. Evaluation of VIE and PIT tagging methods for juvenile cyprinid fishes. Journal of Applied Ichthyology, 25: 381-386.
Brook, B. W., and Bradshaw, C. J. A. 2006. Strength of evidence for density dependence in abundance time series of 1198 species. Ecology, 87: 1445-1451.
Cavrois-Rogacki, T., Davie, A., Monroig, O., and Migaud, H. 2019. Elevated temperature promotes growth and feed efficiency of farmed ballan wrasse juveniles (Labrus bergylta). Aquaculture, 511: 734237. Elsevier. https://doi.org/10.1016/j.aquaculture.2019.734237.
Claudet, J., Pelletier, D., Jouvenel, J. Y., Bachet, F., and Galzin, R. 2006. Assessing the effects of marine protected area (MPA) on a reef fish assemblage in a northwestern Mediterranean marine reserve: Identifying community-based indicators. Biological Conservation, 130: 349-369.
Conover, D. O., and Munch, S. B. 2002. Sustaining fisheries yields over evolutionary time
scales. Science, 297: 94-96.
Costello, M. J. 1991a. Review of the biology of wrasse (Labridae: Pisces) in northern Europe. Progress in Underwater Science, 16: 29-51.
Costello, M. J. 1991b. Review of the biology of wrasse (Labridae: Pisces) in Northern Europe. Science, Progress in Underwater, 16: 29-51.
Costello, M. J. 2006. Ecology of sea lice parasitic on farmed and wild fish. Trends in Parasitology, 22: 475-483.
Cowan, J. H., Rose, K. A., and DeVries, D. R. 2000. Is density-dependent growth in young-of-the-year fishes a question of critical weight? Reviews in Fish Biology and Fisheries, 10: 61-89.
Cushing, D. H. 1974. The possible density-dependence of larval mortality and adult mortality in fishes. In the early life history of fish: 103-111.
Darwall, W. R. T., Costello, M. J., Donnelly, R., and Lysaght, S. 1992. Implications of lifehistory strategies for a new wrasse fishery. Journal of Fish Biology, 41: 111-123.
De Roos, A. M., Boukal, D. S., and Persson, L. 2006. Evolutionary regime shifts in age and size at maturation of exploited fish stocks. Proceedings of the Royal Society B: Biological Sciences, 273: 1873-1880.
Deady, S., and Fives, J. M. 1995. The diet of corkwing wrasse, Crenilabrus melops, in Galway Bay, Ireland, and in Dinard, France. Journal of the Marine Biological Association of the United Kingdom, 75: 635-649.
Dehnhard, N., Langset, M., Aglen, A., Lorentsen, S. H., and Anker-Nilssen, T. 2021. Fish consumption by great cormorants in Norwegian coastal waters - A human-wildlife conflict for wrasses, but not gadids. ICES Journal of Marine Science, 78: 1074-1089.
Doherty, P. J. 1983. Tropical territorial damselfishses : is density limited by aggression or recruitment? Ecology, 64: 176-190.
Edgar, G. J., Stuart-Smith, R. D., Willis, T. J., Kininmonth, S., Baker, S. C., Banks, S., Barrett, N. S., et al. 2014. Global conservation outcomes depend on marine protected areas with five key features. Nature, 506: 216-220.
Espeland, S. H., Nedreaas, K., Mortensen, S., Skiftesvik, A. B., Agnalt, A.-L., Durif, C., Harkestad, L. S., et al. 2010. Kunnskapsstatus leppefisk: Utfordringer i et $\varnothing$ kende fiskeri. Fisken og havet. https://brage.bibsys.no/xmlui/bitstream/handle/11250/113891/FH_72010_til_web.pdf?sequence=1\&isAllowed=y.
Fairbairn, D. J., Blanckenhorn, W. U., and Székely, T. 2007. Sex, size and gender roles: evolutionary studies of sexual size dimorphism. Oxford University Press.
Faust, E., Halvorsen, K. T., Andersen, P., Knutsen, H., and André, C. 2018. Cleaner fish escape salmon farms and hybridize with local wrasse populations. Royal Society Open Science, 5: 171752.
Fenberg, P. B., and Roy, K. 2008. Ecological and evolutionary consequences of size-selective harvesting: How much do we know? Molecular Ecology, 17: 209-220.
Forrester, G. E. 1990. Factors influencing the juvenile demography of a coral reef fish, 71: 1666-1681.
Forskrift om regulering av fisket etter leppefisk i 2022. §§2,4,5,8 (FOR-2021-12-22-3858). https://lovdata.no/dokument/SF/forskrift/2021-12-223858?fbclid=IwAR0iw4dUgNvOlQrXvXJNeuVWWUasSuwP1ODgXScnKQEnkZnXp 7XePVA8u4c (Accessed 13 May 2022).
Frank, K. T., Petrie, B., Choi, J. S., and Leggett, W. C. 2005. Trophic Cascades in a Formerly Cod-Dominated Ecosystem: 1621-1624.
Gårdmark, A., Jonzén, N., and Mangel, M. 1999. Density-dependent processes in structured fish populations: interaction strengths in whole-lake experiments. Ecological Monographs, 69: 155-175.

Gårdmark, A., Jonzén, N., and Mangel, M. 2006. Density-dependent body growth reduces the potential of marine reserves to enhance yields. Journal of Applied Ecology, 43: 61-69.
Gell, F. R., and Roberts, C. M. 2003. Benefits beyond boundaries: The fishery effects of marine reserves. Trends in Ecology and Evolution, 18: 448-455.
Gibbons, J. W., and Andrews, K. M. 2004. PIT tagging: Simple technology at its best. BioScience, 54: 447-454.
Gibson, R. N. 1993. Review of Fishes of the Sea. The North Atlantic and Mediterranean, by J.Lythgoe \& G.Lythgoe. Copeia, 1993: 244-245. https://doi.org/10.2307/1446322.

Gries, G., and Letcher, B. H. 2002. Tag retention and survival of age-0 Atlantic salmon following surgical implantation with passive integrated transponder tags. North American Journal of Fisheries Management, 22: 219-222.
Grorud-Colvert, K., Claudet, J., Tissot, B. N., Caselle, J. E., Carr, M. H., Day, J. C., Friedlander, A. M., et al. 2014. Marine protected area networks: Assessing whether the whole is greater than the sum of its parts. PLoS ONE, 9: e102298.
Gross, M. R. 1982. Sneakers, satellites and parentals: polymorphic mating strategies in North American sunfishes. Zeitschrift für Tierpsychologie, 60: 1-26.
Halpern, B. S., Lester, S. E., and McLeod, K. L. 2010. Placing marine protected areas onto the ecosystem-based management seascape. Proceedings of the National Academy of Sciences of the United States of America, 107: 18312-18317.
Halvorsen, K. T., Sørdalen, T. K., Durif, C., Knutsen, H., Olsen, E. M., Skiftesvik, A. B., Rustand, T. E., et al. 2016. Male-biased sexual size dimorphism in the nest building corkwing wrasse (Symphodus melops): Implications for a size regulated fishery. ICES Journal of Marine Science, 73: 2586-2594.
Halvorsen, K. T., Larsen, T., Sørdalen, T. K., Vøllestad, L. A., Knutsen, H., and Olsen, E. M. 2017a. Impact of harvesting cleaner fish for salmonid aquaculture assessed from replicated coastal marine protected areas. Marine Biology Research, 13: 359-369. Taylor \& Francis. https://doi.org/10.1080/17451000.2016.1262042.
Halvorsen, K. T., Sørdalen, T. K., Vøllestad, L. A., Skiftesvik, A. B., Espeland, S. H., and Olsen, E. M. 2017b. Sex- and size-selective harvesting of corkwing wrasse (Symphodus melops)-a cleaner fish used in salmonid aquaculture. ICES Journal of Marine Science, 74: 660-669.
Halvorsen, K. T., Sørdalen, T. K., Vøllestad, L. A., Skiftesvik, A. B., Espeland, S. H., and Olsen, E. M. 2017c. Sex-and size-selective harvesting of corkwing wrasse (Symphodus melops)-a cleaner fish used in salmonid aquaculture. ICES Journal of Marine Science, 74: 660-669.
Halvorsen, K. T., Sørdalen, T. K., Larsen, T., Browman, H. I., Rafoss, T., Albretsen, J., and Skiftesvik, A. B. 2020. Mind the depth: The vertical dimension of a small-scale coastal fishery shapes selection on species, size, and sex in wrasses. Marine and Coastal Fisheries, 12: 404-422.
Halvorsen, K. T., Skiftesvik, A. B., Durif, C., Faust, E., Wennhage, H., André, C., and Mortensen, S. 2021a. Towards a sustainable fishery and use of cleaner fish in salmonid aquaculture: Challenges and oportunities.
Halvorsen, K. T., Larsen, T., Browman, H. I., Durif, C., Aasen, N., Vøllestad, L. A., Cresci, A., et al. 2021b. Movement patterns of temperate wrasses (Labridae) within a small marine protected area. Journal of Fish Biology, 99: 1513-1518.
Hanson, K. C., Gravel, M. A., Graham, A., Shoji, A., and Cooke, S. J. 2008. Sexual Variation in Fisheries Research and Management: When Does Sex Matter? Reviews in Fisheries Science, 16: 421-436.
Helfman, G., Collette, B. B., Facey, D. E., and Bowen, B. W. 2009. The diversity of fishes: biology, evolution, and ecology. John Wiley \& Sons.

Heuch, P. A., Bjørn, P. A., Finstad, B., Holst, J. C., Asplin, L., and Nilsen, F. 2005. A review of the Norwegian 'National Action Plan Against Salmon Lice on Salmonids': The effect on wild salmonids. Aquaculture, 246: 79-92.
Hilborn, R., Branch, T. A., Ernst, B., Magnussson, A., Minte-Vera, C. V., Scheuerell, M. D., and Valero, J. L. 2003. State of the world's fisheries. Annual Review of Environment and Resources, 28: 359-399.
Hilborn, R. 2018. Are MPAs effective? ICES Journal of Marine Science, 75: 1160-1162.
Hilldén, N. O. 1981. Territoriality and reproductive behaviour in the goldsinny, Ctenolabrus rupestris L. Behavioural Processes, 6: 207-221.
Hixon, M. A., and Webster, M. S. 2002. Density dependence in marine fishes: Coral Reef Populations as model systems BT - Coral reef fishes: dynamics and diversity in a complex ecosystem. Coral reef fishes: dynamics and diversity in a complex ecosystem: 303-325. papers2://publication/uuid/9FAD1BBB-FB27-43D5-919A-65B3509F9023.
Hočevar, S., and Kuparinen, A. 2021. Marine food web perspective to fisheries-induced evolution. Evolutionary Applications, 14: 2378-2391.
Hutchings, J. A., and Baum, J. K. 2005. Measuring marine fish biodiversity: Temporal changes in abundance, life history and demography. Philosophical Transactions of the Royal Society B: Biological Sciences, 360: 315-338.
Jansson, E., Besnier, F., Malde, K., André, C., Dahle, G., and Glover, K. A. 2020. Genome wide analysis reveals genetic divergence between Goldsinny wrasse populations. BMC Genetics, 21: 1-15. BMC Genetics.
Jones, G. P. 1987. Competitive interactions among adults and juveniles in a coral reef fish. Ecology, 68: 1534-1547.
Jones, G. P., Cole, R. C., and Battershill, C. N. 1993. Marine Reserves: Do they work? Proceedings of the Second International Temperate Reef Symposium: 29-45. Auckland. https://www.researchgate.net/publication/261486678.
Jørgensen, C., Enberg, K., Dunlop, E. S., Arlinghaus, R., Boukal, D. S., Brander, K., Ernande, B., et al. 2007. Ecology: Managing evolving fish stocks. Science, 318: 1247-1248.

Kendall, N. W., and Quinn, T. P. 2013. Size-selective fishing affects sex ratios and the opportunity for sexual selection in Alaskan sockeye salmon Oncorhynchus nerka. Oikos, 122: 411-420.
Kimball, M. E., and Mace, M. M. 2020. Survival, growth, and tag retention in estuarine fishes implanted with passive integrated transponder (PIT) tags. Estuaries and Coasts, 43: 151160.

Knutsen, J. A., Kleiven, A. R., Olsen, E. M., Knutsen, H., Espeland, S. H., Sørdalen, T. K., Thorbjørnsen, S. H., et al. 2022. Lobster reserves as a management tool in coastal waters: Two decades of experience in Norway. Marine Policy, 136: 104908.
Korman, J., Yard, M. D., Dzul, M. C., Yackulic, C. B., Dodrill, M. J., Deemer, B. R., and Kennedy, T. A. 2021. Changes in prey, turbidity, and competition reduce somatic growth and cause the collapse of a fish population. Ecological Monographs, 91: e01427.
Kraufvelin, P., Christie, H., and Gitmark, J. K. 2020. Top-down release of mesopredatory fish is a weaker structuring driver of temperate rocky shore communities than bottom-up nutrient enrichment. Marine Biology, 167: 1-20. Springer Berlin Heidelberg. https://doi.org/10.1007/s00227-020-3665-3.
Krkošek, M., Lewis, M. A., Morton, A., Frazer, L. N., and Volpe, J. P. 2006. Epizootics of wild fish induced by farm fish. Proceedings of the National Academy of Sciences of the United States of America, 103: 15506-15510.
Lorenzen, K., and Enberg, K. 2002. Density-dependent growth as a key mechanism in the regulation of fish populations: Evidence from among-population comparisons. Proceedings of the Royal Society B: Biological Sciences, 269: 49-54.

Marshall, D. J., Gaines, S., Warner, R., Barneche, D. R., and Bode, M. 2019. Underestimating the benefits of marine protected areas for the replenishment of fished populations. Frontiers in Ecology and the Environment, 17: 407-413.
McClanahan, T. R., and Mangi, S. 2000. Spillover of exploitable fishes from a marine park and its effect on the adjacent fishery. Ecological Applications, 10: 1792-1805.
McCoy, C. M. R., Dromard, C. R., and Turner, J. R. 2010. An evaluation of Grand Cayman MPA performance : a comparative study of coral reef fish communities.
Moland, E., Fernández-Chacón, A., Sørdalen, T. K., Villegas-Ríos, D., Thorbjørnsen, S. H., Halvorsen, K. T., Huserbråten, M., et al. 2021. Restoration of Abundance and Dynamics of Coastal Fish and Lobster Within Northern Marine Protected Areas Across Two Decades. Frontiers in Marine Science, 8: 1-12.
Nedreaas, K., Aglen, A., Knutsen, H., Smedstad, O., and Ågotnes, P. 2008. Kysttorskforvaltning på Vestlandet og langs Skagerrakkysten: Vurdering av status for kysttorsk på strekningen svenskegrensen-Stad med forslag om forvaltningstiltak.
Nickols, K. J., White, J. W., Malone, D., Carr, M. H., Starr, R. M., Baskett, M. L., Hastings, A., et al. 2019. Setting ecological expectations for adaptive management of marine protected areas. Journal of Applied Ecology, 56: 2376-2385.
Nilsson, P. A., and Brönmark, C. 2000. Prey vulnerability to a gape-size limited predator: behavioural and morphological impacts on northern pike piscivory. Oikos, 88: 539-546.
Norderhaug, K. M., Christie, H., Fosså, J. H., and Fredriksen, S. 2005. Fish-macrofauna interactions in a kelp (Laminaria hyperborea) forest. Journal of the Marine Biological Association of the United Kingdom, 85: 1279-1286.
Norderhaug, K. M., and Moland, E. 2021. Depletion of coastal predatory fish sub-stocks coincided with the largest sea urchin grazing event observed in the NE Atlantic: 163-173.
Nusslé, S., Hendry, A., Knapp, R., Bogan, M., Sturrock, A., and Carlson, S. 2017. Thirty-five experimental fisheries reveal the mechanisms of selection. bioRxiv: 141259. https://www.biorxiv.org/content/10.1101/141259v1\
https://www.biorxiv.org/content /biorxiv/early/2017/05/26/141259.full.pdf.
Overton, K., Dempster, T., Oppedal, F., Kristiansen, T. S., Gismervik, K., and Stien, L. H. 2019. Salmon lice treatments and salmon mortality in Norwegian aquaculture: a review. Reviews in Aquaculture, 11: 1398-1417.
Parenti, P., and Randall, J. E. 2000. an Annotated Checklist of the Species of the Labroid Fish Families Labridae and Scaridae. Ichthyological Bulletin, Number, 68.
Parker, G. A. 1992. The evolution of sexual size dimorphism in fish. Journal of Fish Biology, 41: 1-20.
Pauli, D., Heino, M., Dieckmann, U., and Program, E. 2015. Fisheries-induced Evolution !!! Warning !!! 461-480.
Pauly, D. 1994. From growth to Malthusian overfishing: stages of fisheries resources misuse. Traditional marine resource management and knowledge Information Bulletin.
Perälä, T., and Kuparinen, A. 2020. Eco-evolutionary dynamics driven by fishing: From single species models to dynamic evolution within complex food webs. Evolutionary Applications, 13: 2507-2520.
Persson, L., Andersson, J., Wahlstrom, E., and Eklov, P. 1996. Size-specific interactions in lake Systems : predator gape limitation and prey growth rate and mortality. Ecology, 77: 900911.

Peterson, N. P., Prentice, E. F., and Quinn, T. P. 1994. Comparison of sequential coded wire and passive integrated transponder tags for assessing overwinter growth and survival of juvenile coho salmon. North American Journal of Fisheries Management, 14: 870-873.
Post, J. R., Parkinson, E. A., and Johnston, N. T. 1999. Density-dependent processes in structured fish populations: interaction strengths in whole-lake experiments. Ecological

Monographs, 69: 155-175.
Post, J. R., and Parkinson, E. A. 2001. Energy allocation strategy in young fish: allometry and survival. Ecology, 82: 1040-1051.
Potts, G. W. 1974a. The colouration and its behavioural significance in the corkwing wrasse, crenilabrus melops. Journal of the Marine Biological Association of the United Kingdom, 54: 925-938.
Potts, G. W. 1974b. The colouration and its behavioural significance in the corkwing wrasse, Crenilabrus melops. Journal of the Marine Biological Association of the United Kingdom, 54: 925-938.
Potts, G. W. 1985. The nest structure of the corkwing wrasse, Crenilabrus melops (Labridae:Teleostei). Journal of the Marine Biological Association of the United Kingdom, 65: 531-546.
PRENTICE, and F., E. 1990. Feasibility of using implantable passive integrated transponder (PIT) tags in salmonids. Fish-marking techniques, 7: ages317-322. Am. Fish. Soc. Symp. http://ci.nii.ac.jp/naid/10008427651/en/ (Accessed 28 September 2021).
Pulliam, H. R. 1988. Sources, sinks, and population regulation. The American Naturalist, 132: 652-661.
Quignard, J. P., and Pras, A. 1986. Labridae. Fishes of the North eastern Atlantic and the Mediterranean, 2: 919-942.
R Core Team. 2018. R: A language and environment for statistical computing. R Foundation for Statistical Computing, Vienna, Austria. https://www.r-project.org/ (Accessed 17 May 2022).

Reznick, D. A., Bryga, H., and Endler, J. A. 1990. Experimentally induced life-history evolution in a natural population. Nature, 346: 357-359.
Ricker, W. E. 1954. Stock and recruitment. Journal of the Fisheries Board of Canada, 11:559623.

Robertson, D. R., and Hoffman, S. G. 1977. The roles of female mate choice and predation in the mating systems of some tropical labroid fishes. Zeitschrift für Tierpsychologie, 45: 298-320.
Roos, A. M. De, Boukal, D. S., and Persson, L. 2006. Evolutionary regime shifts in age and size at maturation of exploited fish stocks, 273: 1873-1880.
Rose, K. A., Cowan, J. H., Winemiller, K. O., Myers, R. A., and Hilborn, R. 2001. Compensatory density dependence in fish populations: Importance, controversy, understanding and prognosis. Fish and Fisheries, 2: 293-327.
Rowe, S., and Hutchings, J. A. 2003. Mating systems and the conservation of commercially exploited marine fish. Trends in Ecology and Evolution, 18: 567-572.
Ruud, M. M. 2020. Catchability and selectivity in a multispecies wrasse fishery in Western Norway. (Master`s thesis, University of Agder). http://www.uia.no. Salas, S., Sumaila, U. R., and Pitcher, T. 2004. Short-term decisions of small-scale fishersselecting alternative target species: A choice model. Canadian Journal of Fisheries and Aquatic Sciences, 61: 374-383. Sayer, M. D. ., Gibson, R. N., and Atkinson, R. J. . 1995. Growth, diet and condition of goldsinny on the west coast of Scotland. Journal of Fish Biology, 46: 317-340. Sayer, M. D. J., Gibson, R. N., and Atkinson, R. J. A. 1996. Growth, diet and condition of corkwing wrasse and rock cook on the west coast of Scotland. Journal of Fish Biology, 49: 76-94. Schultz, E. T., and Warner, R. R. 1991. Phenotypic plasticity in life-history traits of female Thalassoma bifasciatum (Pisces: Labridae): 2. Correlation of fecundity and growth rate in comparative studies. Environmental Biology of Fishes, 30: 333-344. Selden, R. L., Gaines, S. D., Hamilton, S. L., and Warner, R. R. 2017. Protection of large predators in a marine reserve alters size-dependent prey mortality. Proceedings of the Royal Society B: Biological Sciences, 284. Shelton, P. A. 2006. Fluctuations and declines in fish populations in the context of species at risk. Sinclair, A. R. E., Mduma, S., and Brashares, J. S. 2003. Patterns of predation in a diverse predator-prey system. Nature, 425: 288-290. Skiftesvik, A. B., Blom, G., Agnalt, A. L., Durif, C. M. F., Browman, H. I., Bjelland, R. M., Harkestad, L. S., et al. 2014. Wrasse (Labridae) as cleaner fish in salmonid aquaculture The Hardangerfjord as a case study. Marine Biology Research, 10: 289-300. Taylor \& Francis. http://dx.doi.org/10.1080/17451000.2013.810760. Skiftesvik, A. B., Durif, C. M. F., Bjelland, R. M., and Browman, H. I. 2015. Distribution and habitat preferences of five species of wrasse (Family Labridae) in a Norwegian fjord. ICES Journal of Marine Science, 72: 890-899. Smokorowski, K. E., and Randall, R. G. 2017. Cautions on using the Before-After-ControlImpact design in environmental effects monitoring programs. FACETS, 2: 212-232. Canadian Science Publishing. Sogard, S. M. 1992. Variability in growth rates of juvenile fishes in different estuarine habitats. Marine Ecology Progress Series, 85: 35-53. Sørdalen, T. K., Halvorsen, K. T., Harrison, H. B., Ellis, C. D., Vøllestad, L. A., Knutsen, H., Moland, E., et al. 2018. Harvesting changes mating behaviour in European lobster. Evolutionary Applications, 11: 963-977. Sparholt, H., Bogstad, B., Christensen, V., Collie, J., Van Gemert, R., Hilborn, R., Horbowy, J., et al. 2021. Estimating Fmsy from an ensemble of data sources to account for density dependence in Northeast Atlantic fish stocks. ICES Journal of Marine Science, 78: 55-69. Stige, L. C., Rogers, L. A., Neuheimer, A. B., Hunsicker, M. E., Yaragina, N. A., Ottersen, G., Ciannelli, L., et al. 2019. Density- and size-dependent mortality in fish early life stages. Fish and Fisheries, 20: 962-976. Sundt, R. C., and Jørstad, K. E. 1998. Genetic population structure of goldsinny wrasse, Ctenolabrus rupestris (L.), in Norway: implications for future management of parasite cleaners in the salmon farming industry. Fisheries Management and Ecology, 5: 291-302. Suski, C. D., Svec, J. H., Ludden, J. B., Phelan, F. J. S., and Philipp, D. P. 2003. The effect of catch-and-release angling on the parental care behavior of male smallmouth bass. Transactions of the American Fisheries Society, 132: 210-218. Sutter, D. A. H., Suski, C. D., Philipp, D. P., Klefoth, T., Wahl, D. H., Kersten, P., Cooke, S. J., et al. 2012. Recreational fishing selectively captures individuals with the highest fitness potential. Proceedings of the National Academy of Sciences of the United States of America, 109: 20960-20965. Svåsand, T., Kristiansen, T. S., Pedersen, T., Salvanes, A. G. V., Engelsen, R., Naevdal, G., and Nødtvedt, M. 2000. The enhancement of cod stocks. Fish and Fisheries, 1: 173-205. Swain, D. P., Sinclair, A. F., and Hanson, J. M. 2007. Evolutionary response to size-selective mortality in an exploited fish population. Proceedings of the Royal Society B: Biological Sciences, 274: 1015-1022. Thangstad, T. 1999. Spatial and temporal distribution of three wrasse species (Pisces : Labridae ) in Masfjord, western Norway: habitat association and effects of environmental variables. Tonn, W., Paszkowski, C., and Holopainen, I. J. 1992. Piscivory and Recruitment : Mechanisms Structuring Prey Populations in Small Lakes. Ecology, 73: 951-958. Treasurer, J. W. 1994. The distribution, age and growth of wrasse (Labridae) in inshore waters of west Scotland. Journal of fish biology, 44: 905-918. Turchin, P. 1995. Population regulation: old arguments and a new synthesis. Population dynamics: new approaches and synthesis: 19-40. Uglem, I., Rosenqvist, G., and Wasslavik, H. S. 2000. Phenotypic variation between dimorphic males in corkwing wrasse. Journal of Fish Biology, 57: 1-14. Uglem, I., and Rosenqvist, G. 2002. Nest building and mating in relation to male size in corkwing wrasse, Symphodus melops. Environmental Biology of Fishes, 63: 17-25. UNCLOS, I. 1982. United Nations convention on the law of the sea. http://www.un.org/Depts/los/convention_agree\%0Aments/convention_overview_conven tion.htm. Uusi-Heikkilä, S., Whiteley, A. R., Kuparinen, A., Matsumura, S., Venturelli, P. A., Wolter, C., Slate, J., et al. 2015a. The evolutionary legacy of size-selective harvesting extends from genes to populations. Evolutionary Applications, 8: 597-620. Uusi-Heikkilä, S., Whiteley, A. R., Kuparinen, A., Matsumura, S., Venturelli, P. A., Wolter, C., Slate, J., et al. 2015b. The evolutionary legacy of size-selective harvesting extends from genes to populations. Evolutionary Applications, 8: 597-620. Wiley-Blackwell. van den Berghe, E. P., and Gross, M. R. 1989. Natural Selection Resulting from Female Breeding Competition in a Pacific Salmon (Coho : Oncorhynchus kisutch). Evolution, 43: 125-140. Victor, B. C. 1986. Larval settlement and juvenile mortality in a recruitment-limited coral reef fish population. Ecological Monographs, 56: 145-160. Vik, Y. K. 2019. Factors affecting growth in corkwing wrasse (Symphodus melops): A study from the west coast of Norway. (Master`s thesis).
Vincenzi, S., Crivelli, A. J., Satterthwaite, W. H., and Mangel, M. 2014. Eco-evolutionary dynamics induced by massive mortality events. Journal of Fish Biology, 85: 8-30.
Walters, C., and Maguire, J. J. 1996. Lessons for stock assessment from the northern cod collapse. Reviews in Fish Biology and Fisheries, 6: 125-137.
Warner, R. R. 1995. Large mating aggregations and daily long-distance spawning migrations in the bluehead wrasse, Thalassoma bifasciatum. Environmental Biology of Fishes, 44: 337-345.
Wickham, H. 2016. gplot2: Elegant Graphics for Data Analysis. Springer-Verlag New York. https://ggplot2.tidyverse.org.
Wood, S. L. R., Jones, S. K., Johnson, J. A., Brauman, K. A., Chaplin-Kramer, R., Fremier, A., Girvetz, E., et al. 2018. Distilling the role of ecosystem services in the Sustainable Development Goals. Ecosystem Services, 29: 70-82. Elsevier B.V. https://doi.org/10.1016/j.ecoser.2017.10.010.
Wright, P. J., and Trippel, E. A. 2009. Fishery-induced demographic changes in the timing of spawning: consequences for reproductive success. Fish and Fisheries, 10: 283-304.
Yuen, J. W., Dempster, T., Oppedal, F., and Hvas, M. 2019. Physiological performance of ballan wrasse (Labrus bergylta) at different temperatures and its implication for cleaner fish usage in salmon aquaculture. Biological Control, 135: 117-123.
Zhou, S., Smith, A. D. M., Punt, A. E., Richardson, A. J., Gibbs, M., Fulton, E. A., Pascoe, S., et al. 2010, May 25. Ecosystem-based fisheries management requires a change to the selective fishing philosophy.
Zuur, A. F., Ieno, E. N., Walker, N. J., Saveliev, A. A., and Smith, G. M. 2009. Mixed effects models and extensions in ecology with R. New York:Springer, 574: 1-4. http://arxiv.org/abs/1305.6995.

## Appendix A

Table A.1: Total number of tagged and recaptured individuals of corkwing and goldsinny over the 5 years of sampling, with numbers from each year instead of each interval.

| Sampling year | Total captures <br> (>100mm) | Tagged <br> individuals | Recap total | Recapture rate <br> (in \%) |
| :---: | :---: | :---: | :---: | :---: |
| Corkwing |  |  |  |  |
| 2017 | 2038 | 1804 | 234 | $11.5 \%$ |
| 2018 | 2852 | 2387 | 465 | $16.3 \%$ |
| 2019 | 1787 | 1399 | 388 | $21.7 \%$ |
| 2020 | 2171 | 1776 | 395 | $18.2 \%$ |
| 2021 | 1897 | 1489 | 408 | $21.5 \%$ |
| Total | 10745 | 8855 | $\mathbf{1 8 9 0}$ | $\mathbf{1 7 . 8} \%$ (mean) |
|  |  |  |  |  |
| Goldsinny |  |  |  |  |
| 2017 | 1285 | 1248 | 37 | 2.9 |
| 2018 | 1511 | 1366 | 145 | 9.6 |
| 2019 | 1094 | 908 | 186 | 17 |
| 2020 | 1142 | 920 | 222 | 19.4 |
| 2021 | 694 | 551 | 143 | 20.6 |
| Total | $\mathbf{5 7 2 6}$ | $\mathbf{4 9 9 3}$ | $\mathbf{7 3 3}$ | $\mathbf{1 3 . 9} \%$ (mean) |

Recapture rate: Using numbers of recaptures in relation to total number of captured individuals larger than 100 mm .

Recap total/total captures) x 100

## Appendix B

Table B.1: Overview of the numerated zone divisions in the control area (Saltskjorholmane) and fished area (Bleikjo). Blue arrows indicate norths direction. Illustration from (Aasen, 2019).


Coordinates for the marine protected area in Austevoll:
N $60^{\circ} 05,514^{\prime} \varnothing 005^{\circ} 16,099$
N $60^{\circ} 05,638^{\prime} \varnothing 005^{\circ} 16,439$
N $60^{\circ} 04,877^{\prime} \emptyset 005^{\circ} 17,809$
N $60^{\circ} 04,736^{\prime} \varnothing 005^{\circ} 17,497$


[^0]:    SUPERVISORS
    Tonje Knutsen Sørdalen, Center for Coastal Research (CCR), University of Agder Kim T. Halvorsen, Institute of Marine Research (IMR)

